

End-labeling of DNA Fragment for DNase I Footprint Analysis

1/28/94 TT

1. Prepare DNA fragment with 3'-recess end (for example, Hind III is OK. Kpn I is not) only at one end.

2.

DNA fragment	100 ng/x μ l
^{32}P - α -dXTP (which can be incorporated into 3' recess end)	2.5 μ l/25 μ Ci
5 mM 3 dNTP	1 μ l x 3 = 3 μ l
10 x Klenow buffer	2.5 μ l
qH ₂ O	(17 - x) μ l /25 μ l
Klenow fragment	0.5 μ l

Incubate at RT for 25 min.

3. Add 1 μ l of 5 mM 4 x dNTP. Incubate at RT 5 min.
4. Add 25 μ l of TE and extract with 100 μ l of P/C/I. Recover the interphase with 40 μ l of TE.
5. Extract with 100 μ l of C/I.
6. Pass the sample through spin column.
7. Add 10 μ l of 3 M NaOAc (pH 5.2) and 300 μ l of 100 % ethanol. Store at -70 °C for 30 min.
8. Spin at 12,000 rpm, 4 °C for 15 min.
9. Wash the pellet twice with 500 μ l of ice-cold 70 % ethanol.
10. Dry the pellet and resuspend in 100 μ l of qH₂O (not in TE).
11. Take 1 μ l and determine cpm. Usually, 1×10^4 cpm/ μ l is expected. Store the labeled probe at 4 °C.

Do not freeze. It is recommended to use within 1 week.