DNA Extraction from Tails
(same protocol good for extracting DNA from any source)

1) Cut tail (about 1 cm) and place in eppendorf tube on dry ice
2) Add 700 µl of tail digestion buffer (see below)
3) Incubate overnight at 50-55C (if necessary, incubation may be as short as a few hours)
4) Vortex the tube and add 700 µl of a 25:24:1 mixture of phenol:chloroform:isoamyl alcohol (PCI) to extract. Mix well and spin 5 min in a microcentrifuge on highest speed.
5) Remove aqueous upper phase to new tube and extract once more as above.
6) Remove aqueous upper phase to new tube and extract with a 24:1 mixture of chloroform:isoamyl alcohol.
7) Transfer 500 µl of the aqueous phase to a fresh tube, add 50 µl of 3M NaOAc, pH 5.2
8) Precipitate by adding 330 µl of isopropanol, mix well.
9) Microcentrifuge for 10 min at highest speed
10) Wash pellet with 500 µl of 70% ETOH, respin.
11) Aspirate remaining liquid and allow samples to air dry under a heat source
12) Resuspend pellets in 50 µl of TE
13) Store at –20C

TAIL DIGESTION BUFFER

6 ml of 0.5 M EDTA

1.5 ml of 1M TRIS, pH 8.0

1.5 ml of 10% SDS

21 ml of QH_2O

30 mg Proteinase K (FISHER, BP1700-100)